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Clinical Manifestations of *Plasmodium berghei* ANKA Infection in Juvenile Mice: A Short Case Report

J. B. Ajayi¹, A. O. Agbeyangi^{1,*}, A. Daniel², I. Omobolaji² and H. O. Mogaji¹

¹Department of Pure and Applied Zoology, Federal University of Agriculture, Abeokuta, Nigeria ²Department of Microbiology, Ogun State Polytechnic, Igbesa, Nigeria *E-mail address: oagbeyangi@yahoo.com

ABSTRACT

Malaria is an important health and development challenge in Africa. Animal models, most particularly mice, have long been employed to study malaria pathogenesis. This paper describes clinical manifestations due to Plasmodium berghei ANKA infection in juvenile mice as a model for understanding the complications of congenital malaria in neonates. Forty-five juvenile mice (5-7 days old) were acquired from University College Hospital, Ibadan and injected with 2×107 (0.2 ml) Plasmodium berghei ANKA parasitized red blood cells (PRBCs). The mice were then transported to the study site, kept in well-ventilated cages and fed daily with a balanced ration. Post-P. berghei infection, the mice were monitored daily for mortality. Clinical manifestations of experimental cerebral malaria (ECM) were assessed and confirmed if at least ruffled fur, hunching, wobbly gait, limb paralysis, convulsions, or coma was observed. Each sign was given a score of 1. Animals with scores \geq 4 were considered to have severe ECM. In the experiment, 20 (44%) mice were lost due to natural cause (i.e. stress) at day 2. Between day 4 and 9, 25 (56%) of the study mice presented clinical signs of ECM. This included: ruffled fur -25 (100%), hunching -21 (84%), wobbly gait -17 (68%), limb paralysis -20(80%), convulsions - 25 (100%). Survival rate and severity of ECM in the mice differs, 22 (88.0%) had severe ECM and 3 (12.0%) had mild ECM. This study has shown that parasite establishment and malaria complications can manifest as early as 4 days' post *P. berghei* infection in 5-7 days old mice.

Keywords: Malaria, Neonates, Juvenile, Mice, ECM, Plasmodium berghei

1. INTRODUCTION

Malaria is a parasitic disease caused by protozoan parasite from the genus *Plasmodium* (*P*.) with female *Anopheles* mosquito acting as the biological vector. Four species of *Plasmodium* are known to infect human, namely *P. ovale, P. vivax, P. malariae* and *P. falciparum*, with the latest being the most virulent and lethal (WHO 2016). Undoubtedly, malaria is still considered as the most serious tropical disease troubling mankind throughout the world with the greatest burden in sub-Saharan African countries. In 2015, there were 214 million cases and 438,000 deaths from this disease, with pregnant women and children under age-five been the most at-risk (WHO, 2015). Children under five are particularly susceptible to malaria illness, infection and death, and about 306,000 of them were killed globally by the disease in 2015 (WHO, 2015).

In non-immune individual, symptoms appear 7 days or more (usually 10-15 days) after the infective mosquito bite. The first symptoms – fever, headache, chills and vomiting – may be mild and difficult to recognize as malaria. If not treated within 24 hours, P. falciparum malaria can progress to severe illness, often leading to death. Children with severe malaria frequently develop one or more of the following symptoms: severe anaemia, respiratory distress in relation to metabolic acidosis, or cerebral malaria. In adults, multi-organ involvement is also frequent (WHO 2015, Fact Sheet. World Malaria Report 2015. Geneva, Switzerland, World Health Organization; WHO, 2015. Fact Sheet. Malaria. Media centre. Geneva, Switzerland, World Health Organization). Experimental animal models have been used in understanding of human disease pathogenesis and development of new drug compounds and vaccines (Drulhe et al., 2002), due to the impossibility of some research procedures on humans for practical or ethical reasons (Basir et al., 2012). So far, Malaria models have been developed in monkey, rats and mice to understand certain aspects of *Plasmodium* pathogenicity in humans. Notably, the mouse model has been extensively studied due to the numerous similarities between the defined malaria antigens in murine and human parasites and also between the murine and human immune response pathway. The many species and strains of Plasmodium available and the very large selection of inbred and outbreed laboratory rodents has created a great number of mouse-parasite combinations that can be used experimentally (Hunt and Grau, 2003; Combes et al., 2005)

In this study, we used *P. berghei* ANKA infection in juvenile mice as a model to investigate the complications of untreated *Plasmodium* infection. We describe clinical manifestations due to *Plasmodium berghei* ANKA infection in juvenile mice as a model for understanding the complications of congenital malaria in neonates.

2. METHODOLOGY

2. 1. Study Area

This study was conducted at the Zoology Laboratory, Department of Pure and Applied Zoology, Federal University of Agriculture, Abeokuta, Ogun State, Nigeria.

2.1.1. Animals

Juvenile mice, 5-7 days old, were used in this study. The mice were obtained from the University College Hospital, Ibadan, and transported to the study site, where they were

maintained at a constant room temperature (25-27 $^{\circ}$ C) in well ventilated cages with unlimited access to food and water. Animals were tagged with a permanent ink conspicuously on the tail for ease of identification.

2. 1. 2. Parasite and infection procedures

Plasmodium berghei ANKA strain (chloroquine sensitive), was obtained from the University College Hospital (UCH), Ibadan. Serial passage of the parasite was initiated by intra-peritoneal (i.p.) injection of 0.2 ml blood containing the parasite into a normal mouse (donor mouse). Subsequent passage for maintaining the parasites were carried out by i.p. inoculation of study mice with 0.2 ml blood, diluted to contain 2×10^7 parasitized red blood cells (PRBC) from the donor mouse to study mice (Franke-Fayard, *et al.* 2004; Ferone *et al.*, 1969; Paton, *et al.*, 1993; Seguin, *et al.*, 1994; Adachi, *et al.*, 2001).

2. 1. 3. Routine observation of infected mice.

Every day after post - *P. berghei* infection, infected mice were routinely monitored for unusual symptoms and eventual mortality (3 times daily). For each of the tagged experimental mouse, clinical symptoms were recorded.

2. 2. Data Analysis

Data obtained were entered using Microsoft Excel 2007 and described using Statistical package for Social Sciences, SPSS Version 20.0 (SPSS Inc. Chicago, Illinois, USA).

3. RESULTS

3. 1. Incidence of death and clinical manifestations in the experimental mice

A total of 45 (100%) mice were monitored over the study period, 20 (44.4%) were lost on day 2 of the experiment. By day 4, 10 (22.2%) had convulsion, and an additional 10 (22.2%) had ruffled fur, hunching, limb paralysis by day 6. On the last day of the experiment, additional 5 (11.1%) of mice had convulsion (**Table 1**).

Days	No of mice lost (%)	Clinical symptoms Before death	Probable cause of death
1	0 (0)	_	NA
2	20 (44.4)	-	Stress
3	-	-	-
4	10 (22.2)	Convulsion	Plasmodium berghei ANKA

Table 1. Incidence of clinical manifestation	Table 1.	Incidence	of clinical	manifestation
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5	-	-	-
6	10 (22.2)	Ruffled fur Hunching limb paralysis Convulsion	Plasmodium berghei ANKA
7	-	-	-
8	-	-	-
9	5 (11.1)	Convulsion	Plasmodium berghei ANKA
Total	45 (100%)		

NA: not applicable

3. 2. Prevalence of Clinical Manifestation in Experimental Mice

Of the 25 mice closely monitored for clinical manifestations, 21 (84.0%) had hunched back, 25 (100%) had ruffled fur, 17 (68.0%) had wobbly gait, 20 (80.0%) had paralyzed limb, and 25 (100%) had convulsion (**Table 2**, **Plates 1**).

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Clinical symptoms	NE	NP (%)
Hunching	25	21 (84.0)
Ruffled fur	25	25 (100.0)
Wobbly gait	25	17 (68.0)
Limb paralysis	25	20 (80.0)
Convulsion	25	25 (100.0)
Coma	25	-

NE: Number examined; NP: Number with clinical presentations



Plates 1

- A- Experimental mice with ruffled fur
- B- Experimental mice with paralyzed limb
- C- Experimental mice with hunched back

3. 3. Prevalence of Experimental Cerebral Malaria (ECM)

For each observed clinical manifestations, a score of one was given to the experimental mice. Animals with scores \geq 4 were considered to have severe ECM and those with lower scores (\leq 3) had mild ECM. Of the 25 (100%) mice observed, 3 (12.0%) had Mild ECM and 22 (88.0%) had severe ECM (**Table 3**).

	NE	NP (%)
Mild ECM		
(ECM = 1, 2, 3)	25	3 (12.0)
Severe ECM		
$(ECM \ge 4)$	25	22 (88.0)

Table 3. Prevalence of Experimental Cerebral Malaria

4. DISCUSSION

Approaches to understanding disease pathogenesis are not new, and have involved experimental infections in animal model and subsequent monitoring of the pathogenic processes (Basir *et al.*, 2012). In this study, we used *P. berghei* ANKA infection in juvenile mice as a model to investigate the complications of untreated *Plasmodium* infection. Data obtained substantiated that juvenile mice from UCH Ibadan are highly susceptible to *P. berghei* ANKA infection, with rapid onset of malaria symptoms beginning on the fourth-day post-inoculation. Clinical manifestations including hunching, ruffled fur, wobbly gait, limb paralysis, convulsions and subsequent death were observed, and are reflective of the high degree of parasitaemia that could be associated with untreated *P. berghei* ANKA infections.

In humans, P. falciparum has been identified as the most virulent and lethal of all Plasmodium species, with various complications and ultimate death. Cerebral Malaria (CM) is one of the major lethal complications of *P. falciparum* infection in human which involved an extremely complex multi-process and multisystem disorder presenting a wide range of clinical features. It is characterized by a sequestration of parasitized red blood cells (PRBC) as the main important feature, particularly in the deep cerebral microvasculature and by the increased levels of pro-inflammatory cytokines (Clarke and Rocket, 1994; Artavanis-Tsakonas et al., 2002). Using the method described by Wu et al. (2010), we identified clinical signs of ECM in our study, with majority of the mice presenting severe ECM before death. Although we could not further investigate the pathogenesis of P. berghei ANKA infection in the mice, this has been adequately described by Basir et al. (2012) in another study. Complications of malaria and subsequent death in mice used during this experiment issues a call for concern in humans, especially neonates exposed to *Plasmodium* parasites and left untreated. It is therefore important to intensify on efforts that prevent congenital transmission of malaria parasite between mother and child, and as well be keen about procedures of prompt diagnosis and treatment of malaria disease in children with developing immunity.

5. CONCLUSION

Clinical manifestations observed during this study are reflective of the high degree of parasitaemia that could be associated with untreated *P. berghei* ANKA infections. Such complications issue a call for concern in humans, especially neonates exposed to *Plasmodium* parasites and left untreated. It is therefore important to intensify the efforts that prevent congenital transmission of malaria parasite between mother and child, and as well be keen about procedures of prompt diagnosis and treatment of malaria disease in children with developing immunity.

References

- [1] Druilhe P., Hagan P., Rook G.A.W. (2002). The importance of models of infection in the studyof disease resistance. *Trends Microbiol*. 10(10), 38-46
- [2] Basi, R., Fazalul Rahiman, S.S., Hasballah, K., Chong, W.C., Talib, H., Yam, M.F., Jabbarzare, M., Tie, T.H., Othman, F., Moklas, M.M., Abdullah, W.O., and Ahmad Z.

(2012). Plasmodium berghei ANKA Infection in ICR Mice as a Model of Cerebral Malaria. *Iranian J Parasitol*. 7(4), 62-74

- [3] Hunt N.H., Grau G.E. (2003). Cytokines: accelerators and brakes in the pathogenesis of cerebral malaria. *Trends Immunol*. 24(9), 491-499.
- [4] Combes V., de Souza J.B., Renia L., Hunt N.H., Grau G.E. (2005). Cerebral malaria: which parasite? Which model? *Drug Disc Today* 2(2), 141-147
- [5] Clark, I.A. and Rockett K.A. (1994). The cytokine theory of human cerebral malaria. *Parasitol Today* 10(10), 410-412
- [6] Artavanis-Tsakonas K. and Riley E.M. (2002). Innate immune response to malaria: rapid induction of IFNγ from human NK cells by live Plasmodium falciparum-infected erythrocytes. J. Immunol. 169(6), 2956-2963
- [7] Wu, J.J., Chen, G., Liu, J., Wang, T., Zheng, W., and Co, Y.M. (2010). Natural regulatory T cells mediate the development of cerebral malaria by modifying the proinflammatory response. *Parasitol Int.* 59, 232–241
- [8] Blandine Franke-Fayard, Holly Trueman, Jai Ramesar, Jacqui Mendoza, Maartenvan der Keur, Reiniervan der Linden, Robert E. Sinden, Andrew P. Waters, Chris J. Janse. A *Plasmodium berghei* reference line that constitutively expresses GFP at a high level throughout the complete life cycle. *Molecular and Biochemical Parasitology*, Volume 137, Issue 1, September 2004, Pages 23-33.
- [9] R. Ferone, J.J. Burchall, and G.H. Hitchings. *Plasmodium berghei* Dihydrofolate Reductase Isolation, Properties, and Inhibition by Antifolates. *Molecular Pharmacology January* 1969, 5 (1) 49-59
- [10] Michael G. Paton, Guy C. Barker, Hiroyuki Matsuoka, Jai Ramesar, Chris J. Janse, Andy P. Waters, Robert E. Sinden. Structure and expression of a post-transcriptionally regulated malaria gene encoding a surface protein from the sexual stages of *Plasmodium berghei*. *Molecular and Biochemical Parasitology*, Volume 59, Issue 2, June 1993, Pages 263-275
- [11] M C. Seguin, F. W. Klotz, I. Schneider, J.P. Weir, M. Goodbary, M. Slayter, J.J. Raney, J.U. Aniagolu, S.J. Green. Induction of nitric oxide synthase protects against malaria in mice exposed to irradiated Plasmodium berghei infected mosquitoes: involvement of interferon gamma and CD8+ T cells. *The Journal of Experimental Medicine* 1994, 1 July, 180 (1), 353
- [12] Keishi Adachi, Hiroko Tsutsui, Shin-Ichiro Kashiwamura, Ekihiro Seki, Hiroki Nakano, Osamu Takeuchi, Kazuyoshi Takeda, Ko Okumura, Luc Van Kaer, Haruki Okamura, Shizuo Akira and Kenji Nakanishi. *Plasmodium berghei* Infection in Mice Induces Liver Injury by an IL-12- and Toll-Like Receptor/Myeloid Differentiation Factor 88-Dependent Mechanism. *J Immunol*. November 15, 2001, 167 (10), 5928-5934